SYSTEMATIC REVIEW

Predicting SARS-Cov-2 - Procoagulant/Coagulation Related Protein Interaction: A Systematic Review

Tonang Dwi Ardyanto¹, Rasmaya Niruri², Adi Yugatama², Artini Pangastuti³

¹ Department of Clinical Pathology, Faculty of Medicine, Universitas Sebelas Maret, 57126, Surakarta, Indonesia,

² Department of Pharmacy, Faculty of Math and Science, Universitas Sebelas Maret, 57126 Indonesia.

³ Department of Biology, Faculty of Math and Science, Universitas Sebelas Maret, 57126 Indonesia.

ABSTRACT

Introduction: Patients with COVID-19 tend to have procoagulant states and a higher risk of blood clotting. This systematic review aimed to provide an overview of studies that used the computational docking method to predict the interaction between procoagulant/coagulation human proteins and SARS-CoV-2. Method: A systematic review was conducted using six databases (Scopus, Cochrane, PubMed, Clinical key, Emerald, and Google scholar). The review included original articles, that applied computational docking methods for viewing the potential interaction between host protein and SARS-CoV-2 in COVID-associated coagulopathy. Results: Five of 715 articles were included. In this systematic review, the assessment of high-quality aspects included multi-target protein assessment, the utilization of crystal structure for the generation of target interaction, the use of CAPRI validated protein-protein docking tools combined with other software, molecular dynamic simulation, and multi-host protein databases. The systematic review revealed the prediction interaction between host procoagulant/coagulation related protein and SARS-CoV-2 protein as follows: VKORC1 - ORF7a, Thrombin - Protein S, ORF3a-heme. Protein S has three potential Heme-binding motifs (HBMs) and protein 7a contains two HBMs. Thrombin and Factor Xa were structurally similar to Mpro. Human gene variants influence the affinity interaction between SARS-CoV-2 and human proteins. Discussion: This systematic review provided a list of SARS-CoV-2 protein targets (ORF7a, Protein S, ORF3a, Mpro) that had the potential to interact with human proteins through different mechanisms of action of procoagulant state and blood clotting. This will expand the diagnostic, clinical monitoring, and therapeutic options for SARS-CoV-2 infection.

Keywords: Computation; Molecular; COVID-19; Blood Coagulation

Corresponding Author:

Rasmaya Niruri, S.Si. M.Fram. Klin. Apt Email: rasmaya@staff.uns.ac.id Tel: +62 81548552828

INTRODUCTION

Coronavirus disease (COVID-19) is caused by severe acute respiratory syndrome coronavirus (SARS-CoV-2). This viral infection has caused a global pandemic. It has spread to more than 200 countries worldwide. Patients with COVID-19 infections may tend to have procoagulant states and a higher risk of blood clots. Coagulation disorders are challenging to treat. This is an important feature of disease severity (1,2). Autopsies identified clots in vessels of COVID-19 patients' lungs, liver, heart, and kidney. Excessive activation of platelets and coagulation proteins may cause thrombocytopenia and high levels of blood clots. SARS-CoV-2 infection evokes a systemic inflammatory response and causes procoagulant-anticoagulant homeostatic abnormalities (1–4). Heme may also increase procoagulant activity and induce coagulation activation. Hemolytic anemia has been reported in patients with severe COVID-19. In this pathological state, free heme is released from hemoglobin. It promotes coagulation activation (5–7).

Based on a recent systematic review-meta-analysis study, COVID-19 coagulopathy and its complications have been considered to correlate with severe disease and a higher risk of death. Thromboembolism was significantly associated with the COVID-19 mortality rate. The mechanism of SARS-CoV-2 protein interactions with human procoagulant/coagulopathy proteins is still being understood (8). Improving our understanding of SARS-CoV-2 infections will expand the diagnostic, patient care, and therapeutic options. A computational approach is a time-saving method with the best chance of identifying potential interactions between SARS-CoV-2 and the host protein. This systematic review aimed to provide an overview of studies that have used computational approaches to predict the interaction between human procoagulant/ coagulation-related proteins and SARS-CoV-2.

MATERIALS AND METHODS

The procedure, search strategy, and study selection The protocol of Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) was applied in this systematic review. Problem (P), Intervention (I), Control (C), and Outcome (O) in this study were COVID-19 (P), SARS-CoV-2 protein/gene(I), human procoagulant/coagulopathy related protein/gene(C), and computational approaches for protein-protein interaction(O). We searched in six databases (Scopus, Cochrane, PubMed, Clinical key, Emerald, and Google scholar) with keywords ("DOCKING") AND ("GENE" OR "PROTEIN") AND ("COAGULATION") AND ("COVID"). Our systematic review included original studies (published since 2020) that used computational approaches for SARS-CoV-2–Procoagulant/coagulation protein interactions. Studies with a wet laboratory apt. Rasmaya Niruri, S.Si. M.Fram. Klin. Apt. approach, non-English language articles, duplicate articles, and the inaccessibility of full-text were excluded. Two authors independently conducted article screening, reviewed articles, and assessed the quality of Universitas Sebelas Maret articles. In the case of discrepancy results, a third author was involved for a consultation to reach a consensus.

Quality assessment

Based on potential issues of bias in the computational protein-ligand study, Mohamed proposed five aspects (Mohamed tools) for assessing quality in a systematic review (9). We used Mohamed tools to assess the study quality in our systematic review. To identify protein-protein interactions, we modified two items of the Mohammed tools, which were docking tools and resources of databases. In Mohamed-tools "the use of AutoDock-Vina combined with other molecular docking-tools" was considered as one of the criteria of a high-quality study(9). Compared to AutoDock-Vina, Autodock tool had a similar performance. However, big ligand analysis remains a challenge for these two docking tools (10). Therefore, we changed the option with Critical Assessment of Predicted Interaction (CAPRI) validated protein-protein docking tools (11). Furthermore, we also changed "Mohamed's item "resources for approved drugs" (9) into "resources for host protein". Multiple databases give a better view of the structure and function of proteins (12). Therefore, in this systematic review, the assessment of high-quality aspects included multi-target protein analysis, the utilization of crystal structure for assessing the target interaction, the use of CAPRI validated protein-protein docking tools combined with other software, multi-host protein databases, and molecular dynamic simulation.

RESULTS

Study Selection

Five of 715 articles were included in this systematic review (Figure 1). Methods/protocol, software, protein target, and human procoagulant/coagulation protein

TYPE OF MANUSCRIPT : Systematic Review

Predicting SARS-Cov-2 - Procoagulant/Coagulation Related Protein Interaction:

A Systematic Review

Tonang Dwi Ardyanto¹, Rasmaya Niruri^{2*}, Adi Yugatama², Artini Pangastuti³

¹ Department of Clinical Pathology, Faculty of Medicine, Universitas Sebelas Maret, 57126,

Surakarta, Indonesia,

² Department of Pharmacy, Faculty of Math and Science, Universitas Sebelas Maret,

Indonesia, Postal address: 57126

³ Department of Biology, Faculty of Math and Science, Universitas Sebelas Maret, Indonesia, Postal address: 57126

*Corresponding Author

Department of Pharmacy

Faculty Math and Science

Jalan IR Sutami 36 A. Surakarta. Indonesia. 67126 Figure 1: Flow diagram of the systematic review process following PRISMA

varied among the included studies (Table I and II).

Study Quality

The result of the quality assessment were in Figure 2 and Table I. The total number of high-quality aspects was 4 (1 article), 3 (1 article), 2 (2 articles), and 1 (1 article). None of the studies performed molecular dynamic simulation.

Data Synthesis

Potential protein target candidates of SARS-CoV-2 were Protein S, ORF3a, ORF7a, and Mpro (13-17). This systematic review revealed the best prediction interaction between host procoagulant/coagulationsrelated protein and SARS-CoV-2 protein as follows: VKORC1-ORF7a, Thrombin-SER494 in protein S, and ORF3a-heme (Table II).

DISCUSSION

The potential SARS-CoV-2 proteins identified in this study were protein S, ORF3a, ORF 7a, and Mpro (Table 2). Protein S attaches to the host angiotensin-converting enzyme 2 (ACE2). Protein S is essential for receptor recognition. ORF3a is a protein that contributes to activating NLRP3, ASC ubiquitination, caspase 1, and IL-1β maturation. ORF7a is a type I transmembrane protein (18,19). ORF7a binds to human CD14+ monocytes, leading to decreased HLA-DR/DP/ DQ levels and increase proinflammatory cytokine levels (20). Mpro is essential for SARS-CoV-2 gene transcription, viral ribonucleic acid translation, and viral replication(21).

Design of target proteins		Target template Modelling			Docking Tools			Mol dynam la	ecular nic simu- tion	Resources of host protein		Total high- quality aspects	References
1	<u>2</u>	<u>3</u>	4	5	6	<u>7</u>	8	<u>9</u>	10	11	<u>12</u>		
	V	V			V				V	NR		2	(13)
	V	V	V			V	V		V		BIOGRID, NCBI	4	(14)
	V	V	V				V		V	RCSB PDP		2	(15)
	V	V	V				V		V		NCBI, RSCB PDB	3	(16)
V		V			V				V	RCSB PDP		1	(17)
1=Mono-targ	get; 2=Multi-ta	arget; 3=Cr	ystal structu	ure; 4=Hom	ology model	ling; 5=Co-cr	ystal ligar	nd; 6=Autod	oct-Vina/Auto	odock; 7=CAPRI va	lidated protein-prot	ein docking-to	ols combined with

Table I: Quality of studies included in the systematic review

1=Mono-target; 2=Multi-target; 3=Crystal structure; 4=Homology modelling; 5=Co-crystal ligand; 6=Autodoct-Vina/Autodock; /=CAPRI validated protein-protein docking-tools combined with other software; 8=Others; 9=Yes; 10=No; 11=mono-resource; 12=multi-resources; NR=Not reported; RSCB PDB=RSCB Protein Bank Database; BIOGRID=the biological general repository for interaction datasets; NCBI=national center for biotechnology information; <u>underlined text</u>=criteria of high quality aspect (which was number 2,3,7,9,12).

Table II: Human Procoagulant/ Coagulation Proteins Interaction with SARS Cov-2 Proteins

Human protein references	O R F 10	O R F 7a	O R F 3a	O R F 1ab	N	S	М	E	M pro	Main findings	
Thrombin (13)						V				Thrombin links with Protein S and also binds with ACE2.	
VKORC1 (14)		V								VKORC1-ORF7a possesses a strong binding affinity. VCORC1 gene vari- ants might influence this interaction.	
SERPING 1 (14)		Х									
PABPC 4 (14)		Х									
Heme (15)		V				V	х	х		Protein S is a moderate binder and ORF7a is a poor.	
Heme (16)	V		V							Heme-iron binding site was Arg134 of ORF3a, Ile304 of N, and Cys44 of	
Human-Oxy-Hemoglobin (16)	V		V	V	V					E. The important interactions were ORF3a-cytochrome c (G41S), and	
Human-Deoxy- Hemoglobin (16)	V		V	V	V					oxidized hemoglobin/ deoxyhemoglo- bin - ORF1ab-ORF3a-ORF10.	
Cytochrome c (G41S) (16)			V								
Human erythrocyte catalase (16)			Х								
Human Erythrocyte NADH-cyto- chrome b5 Reductases(16)			Х								
Factor Xa (17)									V	Similarities in structural comparison	
Thrombin (17)									V	while the number allowing Mpto and the man protein (Factor Xa and thrombin), which were presented by low root mean-square deviation values (2,49 Å and 2.57Å).	

V=potential candidate; X=dismissed as candidate because of lack of structural data or no similar motif.

ORF 7a-human VKORC1 possessed a strong binding affinity (Table 2). VKORC1 plays an essential role in supporting coagulation activity through the vitamin K cycle (22). Gene variants of VKORC1 may contribute to the binding mechanism. The gene variants may affect splicing, minimum free energy (MFE) value of messenger ribonucleic acid (mRNA), and micro ribonucleic acid

(miRNA). Splicing alterations were found at -695C T, +1391 C>T, and +2110T>C. The change in splicing may affect mRNA and alter potential binding affinity to the virus. Furthermore, a potential increase in mRNA MFE value was identified in VKORC -1975G>C and +1100 C>T. A greater MFE value may give greater feasibility for mRNA degradation. Changes were also determined using miRNA analysis. In this case, miRNA increases in -2834C>A and -1675G>A, and miRNA decreases in -3537C>G. An elevation in miRNA may lower protein expression, and vice versa. The interaction between VKORC1 and ORF7a may inhibit the activation of clotting proteins (14). This may have affect the substances metabolized by VKORC1. The anticoagulant warfarin, inhibits the Vitamin K-epoxide reductase complex. Warfarin is also metabolized in CYP2C9 (a part of the CYPP450 family). VKORC1 and CYP2C9 are related to warfarin sensitivity and dose requirement (23,24). The significant features of symptomatic COVID-19 were cytokine release, proinflammatory development, and lower CYP450 isoenzyme levels. These features may decrease warfarin metabolism (25-27). Decreasing warfarin dose requirements may be required for patients with COVID-19 (25). Interestingly, patients with COVID-19 tend to have a lower vitamin K status as compared to healthy subjects (28). It is essential to understand the effect of SARS-CoV-2 infection on warfarin dose adjustment.

Another interaction between SARS-Cov-2 and the host protein is Protein S-thrombin. The docking result showed that thrombin is linked with Ser494 in Protein S and also Glu37 in human receptor ACE2 (13). SARS-Cov2 protein S (Ser494, Phe 486, Leu 455) supports hotspot 31 of the ACE2 receptor. Ser494 also strengthenened the stability of hotspot 353 (29,30). In vitro tests, using cell-based bioassays, showed that human protease (thrombin and factor Xa) can promote viral entry by direct proteolytic cleavage of SARS-CoV-2 Protein S (31). Factor X is vitamin K dependent. It is expressed mainly in the liver and in other cells which express ACE-2. Factor X, a localized serine protease, cleaves protein S, which binds to the host cell receptor. Spike protein 2 is released. This supports the fusion of the host cell membrane and Sars-CoV-2. The Sars-CoV-2 genetic material was inserted into the human cells. It replicates in the host cells. New Sars-CoV-2 particles are released and cause infection of other host cells, inflammation, and coagulation (32). Therefore, anticoagulation therapy may suppress viral entry (31).

Thrombin bursts have also been reported in cases of severe COVID-19 (33). Sars-Cov-2 may also affect coagulation through Mpro (17). Thrombin is highly similar to the active site of Mpro, leading to the potential for off-target binding of small molecules (34). Human proteases (Factor Xa and thrombin) had fold similarity with SARS-CoV-2 Mpro, which were presented by low root-mean-square deviation values (2,49 Å and 2.57Å). These superposed structures may cause perturbations in the coagulation (17). Disorder in pro-clotting factors caused a significantly lower value of antithrombin and a higher D-dimer level and fibrinogen. The higher value of D-dimer showed worse progression of COVID-19 severity due to disseminated intravascular coagulation or thrombosis. These features may be caused by

activation of the coagulation system (4,17,35). Blocking coagulation proteins are potential targets for anticoagulant therapy in COVID-19 (34).

SARS-Cov-2 proteins (Protein S and ORF3a) are also linked to Heme. Direct interactions of SARS-Cov-2 proteins with heme are presented by Heme Binding Motifs (HBMs). Protein S has three HBMs, a positive net charge (+2), and several potential hydrophobic residues that moderately support heme binding. ORF7a possesses only two hydrophobic residues that support heme binding. ORF7a is a poor heme binder with two HBMs. Therefore, Protein S becomes a more interesting candidate than OFR7a for interaction with heme (15). Heme is also essential for Protein S to avoid antibody immunity. Free heme is toxic. An endogenous tetrapyrrole product, biliverdin, is produced for heme detoxification. High-affinity binding of N-terminal domain (NTD) of Protein S to biliverdin stabilizes the NTD structure and produce the SARS-CoV-2 envelope glycoprotein. This leads to reduces the reactivity of Protein S and blocks the neutralizing antibody response to the virus (36).

Heme is composed of protoporphyrin IX and iron. Heme binds to hemoglobin and cytochrome c (Cyt c) to play crucial roles in human biological processes (37). Dysregulation of heme-iron metabolism and hemoglobin dysfunction has been identified in COVID-19 (38). The heme-iron binding sites identified in the conserved-domain analysis, were Arg134 of ORF3a, Ile304 of protein N, and Cys44 of protein E. The conserved domains of ORF3a were also found in Cyt c. ORF3a does not interact with wild-type Cyt c, but it fully embeds in Cyt c G41S variant and blocks heme. The interaction of ORF3a-Cyt c (G41S) may contribute to the respiratory problems in COVID-19. Based on this result, subjects with Cyt c (G41S) may have a greater risk of disease severity. The docking results also showed that ORF1ab, ORF3a, and ORF10 were bound to oxidized hemoglobin and deoxyhemoglobin. ORF3a and ORF10 of SARS Cov-2 are embeded in deoxyhemoglobin and directly bind to the heme of the beta chain. This result may explain the invasion of the virus on hemoglobin (16). CD26, CD147, and ACE2, which are located on erythrocytes and blood cell precursors, also play important roles in the interaction between hemoglobin molecules and SARS Cov-2 (38). SARS-Cov-2 induces hemoglobin and heme-iron dysfunction. Consequently, it may cause anemia, hyperferritinemia, an increase in free toxic heme in circulation, downregulation of heme oxygenase-1, hypoxemia, hypoxia, an increase in oxidative stress, inflammatory stress, and multiorgan damage (38-40). The incidence of anemia is not frequent in patients infected with SARS-CoV-2. Low levels of hemoglobin have been identified in severe disease (6,7,41). Free heme increases oxidative stress by scavenging nitric oxide, catalyzing the Fenton reaction, and activating inflammasome (40). Excess-free heme in circulation may induce thrombin generation (5).

The COVID-19 pandemic is a challenging phenomenon in the healthcare system (42). During this pandemic, hospitals experienced a large number of COVID-19 patients. Interprofessional collaboration was adopted to promote patient care. COVID 19 can cause many clinical manifestations and complications in multiple organ systems. Therefore, a multi-professional team is involved in COVID-19 management (43). The SARS-CoV-2 infection causes coagulation disorders. Patients with COVID coagulopathy have a higher incidence in severe cases and have a poor prognostis (44). The health care team is a vulnerable group that is not only exposed to the hazards of the pathogen but also psychological stress. Updating knowledge of SARS CoV-2 is important for health services (45,46). Our systematic review results can be beneficial for future clinical investigations.

All computational docking methods have inherent limitations to describe protein interactions models. Therefore, investigation in wet laboratories and clinical studies need to be conducted for further exploration.

CONCLUSION

This systematic review provided a list of SARS CoV-2 protein targets (ORF7a, Protein S, ORF3a) that had the potential to interact with human protein (VKORC-1, thrombin, and heme) through different mechanisms of action of procoagulant state and blood clotting. Therefore, it will expand the diagnostic, clinical monitoring, and therapeutic options in SARS-CoV-2 infection.

REFERENCES

- Biswas S, Thakur V, Kaur P, Khan A, Kulshrestha S, Kumar P. Blood clots in COVID-19 patients: Simplifying the curious mystery. Med Hypotheses. 2021 Jan 1;146. https://dx.doi.org/10.1016%2Fj. mehy.2020.110371
- 2. Giannis D, Ziogas IA, Gianni P. Coagulation disorders in coronavirus infected patients: COVID-19, SARS-CoV-1, MERS-CoV and lessons from the past. J Clin Virol. 2020 Jun 1;127. https:// doi.org/10.1016/j.jcv.2020.104362
- 3. Levi M, Thachil J, Iba T, Levy JH. Coagulation abnormalities and thrombosis in patients with COVID-19. Vol. 7, The Lancet Haematology. Elsevier Ltd; 2020. p. e438–40. https://doi. org/10.1016/s2352-3026(20)30145-9
- 4. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. J Thromb Haemost. 2020 Apr 1;18(4):844–7. https:// doi.org/10.1111/jth.14768
- 5. Sparkenbaugh EM, Chantrathammachart P, Wang S, Jonas W, Kirchhofer D, Gailani D, et al.

Excess of heme induces tissue factor-dependent activation of coagulation in mice. Haematologica. 2015;100(3):308–13. https://doi.org/10.3324/ haematol.2014.114728

- Mokhtari T, Hassani F, Ghaffari N, Ebrahimi B, Yarahmadi A, Hassanzadeh G. COVID-19 and multiorgan failure: A narrative review on potential mechanisms. Vol. 51, Journal of Molecular Histology. Springer Science and Business Media B.V.; 2020. p. 613–28. https://doi.org/10.1007/ s10735-020-09915-3
- Sahu KK, Borogovac A, Cerny J. COVID-19 related immune hemolysis and thrombocytopenia. J Med Virol. 2021 Feb 1;93(2):1164–70. https://doi. org/10.1002/jmv.26402
- 8. Malas MB, Naazie IN, Elsayed N, Mathlouthi A, Marmor R, Clary B. Thromboembolism risk of COVID-19 is high and associated with a higher risk of mortality: A systematic review and metaanalysis. EClinicalMedicine. 2020 Dec 1;29–30. https://doi.org/10.1016/j.eclinm.2020.100639
- 9. Mohamed K, Yazdanpanah N, Saghazadeh A, Rezaei N. Computational drug discovery and repurposing for the treatment of COVID-19: a systematic review. Bioorganic Chem . 2021 Jan;106:104490. https://doi.org/10.1016/j. bioorg.2020.104490
- 10. Vieira TF, Sousa SF. Comparing AutoDock and Vina in ligand/decoy discrimination for virtual screening. Appl Sci. 2019 Nov 1;9(21). https://doi. org/10.3390/app9214538
- 11. Kangueane P, Nilofer C. Protein-Protein Docking: Methods and Tools. In: Protein-Protein and Domain-Domain Interactions. Springer Singapore; 2018. p. 161–8. DOI: 10.1007/978-981-10-7347-2_14-7347-2_14
- 12. Xu D. Protein databases on the Internet. Curr Protoc Mol Biol. 2012 Jan;Chapter 19:Unit 19.4. https://doi.org/10.1002/0471142727.mb1904s97
- 13. Bhanu P, Kumar NH, Hemandhar Kumar S, Relekar M, Anand DA, Kumar J. Comparative molecular docking analysis of the SARS CoV-2 Spike glycoprotein with the human ACE-2 receptors and thrombin. Bioinformation. 2020 Jul 31;16(7):532–8. https://doi.org/10.6026/97320630016532
- 14. Holcomb D, Alexaki A, Hernandez N, Laurie K, Kames J, Hamasaki-Katagiri N, et al. Potential impact on coagulopathy of gene variants of coagulation related proteins that interact with SARS-CoV-2. bioRxiv 2020 Sep 18:2020.09.08.272328. doi: 10.1101/2020.09.08.272328
- Hopp MT, Domingo-Fern6ndez D, Gadiya Y, Detzel MS, Schmalohr BF, Steinbock F, et al. Unravelling the debate on heme effects in COVID-19 infections. bioRxiv. 2020; https://doi. org/10.1101/2020.06.09.142125
- 16. Liu W LH. COVID 19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism. ChemRxiv

Cambridge Cambridge Open Engag. 2020; https://doi.org/10.26434/chemrxiv.11938173.v7

- 17. Biembengut HV, de Souza T de ACB. Coagulation modifiers targeting sars-cov-2 main protease mpro for covid-19 treatment: An in silico approach. Mem Inst Oswaldo Cruz. 2020 May 1;115(5):1–4. https://doi.org/10.1590/0074-02760200179
- Yoshimoto FK. The proteins of severe acute respiratory syndrome coronavirus-2 (SARS CoV-2 or n-COV19), the cause of COVID-19. Protein J. 2020 Jun;39:198–216. https://doi.org/10.1007/ s10930-020-09901-4
- 19. Huang Y, Yang C, Xu X feng, Xu W, Liu S wen. Structural and functional properties of SARS-CoV-2 spike protein: potential antivirus drug development for COVID-19. Vol. 41, Acta Pharmacologica Sinica. Springer Nature; 2020. p. 1141–9. https:// doi.org/10.1038/s41401-020-0485-4
- 20. Zhou Z, Huang C, Zhou Z, Huang Z, Su L, Kang S, et al. Structural insight reveals SARS-CoV-2 ORF7a as an immunomodulating factor for human CD14+ monocytes. iScience. 2021 Mar 19;24(3). https:// doi.org/10.1016/j.isci.2021.102187
- Trougakos IP, Stamatelopoulos K, Terpos E, Tsitsilonis OE, Aivalioti E, Paraskevis D, et al. Insights to SARS-CoV-2 life cycle, pathophysiology, and rationalized treatments that target COVID-19 clinical complications. Vol. 28, Journal of Biomedical Science. BioMed Central Ltd; 2021. 28(1):1-18. https://doi.org/10.1186/s12929-020-00703-5
- 22. Li S, Liu S, Liu XR, Zhang MM, Li W. Competitive tight-binding inhibition of VKORC1 underlies warfarin dosage variation and antidotal efficacy. Blood Adv. 2020 May 26;4(10):2202–12. https://doi.org/10.1182/bloodadvances.2020001750
- 23. Ahmed S, Altaf N, Ejaz M, Altaf A, Amin A, Janjua K, et al. Variations in the frequencies of polymorphisms in the CYP2C9 gene in six major ethnicities of Pakistan. Sci Rep. 2020 Dec 1;10(1). https://doi.org/10.1038/s41598-020-76366-x
- 24. Al Ammari M, AlBalwi M, Sultana K, Alabdulkareem IB, Almuzzaini B, Almakhlafi NS, et al. The effect of the VKORC1 promoter variant on warfarin responsiveness in the Saudi WArfarin Pharmacogenetic (SWAP) cohort. Sci Rep. 2020 Dec 1;10(1). https://doi.org/10.1038/s41598-020-68519-9
- 25. Irwin MN, Adie S, Sandison K, Alsomairy SA, Brancaccio A. Warfarin Dose Requirements in Adults Hospitalized With COVID-19 Infection: A Retrospective Case Series. J Pharm Pract. 2021; Mar 15:08971900211000705. https://doi. org/10.1177/08971900211000705
- 26. Tay MZ, Poh CM, Rŭnia L, MacAry PA, Ng LFP. The trinity of COVID-19: immunity, inflammation and intervention. Nat Rev Immunol. 2020 Jun;20(6):363–74. https://doi.org/10.1038/s41577-020-0311-8

- Bleau AM, Levitchi MC, Maurice H, du Souich P. Cytochrome P450 inactivation by serum from humans with a viral infection and serum from rabbits with a turpentine-induced inflammation: The role of cytokines. Br J Pharmacol. 2000 Aug; 130: 1777 – 84. https://doi.org/10.1038/ sj.bjp.0703486
- 28. Linneberg A, Kampmann FB, Israelsen SB, Andersen LR, Jurgensen HL, Sandholt H, et al. The association of low vitamin k status with mortality in a cohort of 138 hospitalized patients with covid-19. Nutrients. 2021 Jun 1;13(6). https://doi. org/10.3390/nu13061985
- 29. Choudhary S, Malik YS, Tomar S. Identification of SARS-CoV-2 Cell Entry Inhibitors by Drug Repurposing Using in silico Structure-Based Virtual Screening Approach. Front Immunol. 2020 Jul 10;11. https://doi.org/10.3389/fimmu.2020.01664
- 30. Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus. J Virol. 2020 Mar 17;94(7). https://doi.org/10.1128/ JVI.00127-20
- 31. Kastenhuber ER, Jaimes JA, Johnson JL, Mercadante M, Muecksch F, Weisblum Y, et al. Coagulation factors directly cleave SARS-CoV-2 spike and enhance viral entry. bioRxiv. 2021 Apr 1:2021.03.31.437960. doi: 10.1101/2021.03.31.437960.
- 32. Frydman GH, Streiff MB, Connors JM, Piazza G. The Potential Role of Coagulation Factor Xa in the Pathophysiology of COVID-19: A Role for Anticoagulants as Multimodal Therapeutic Agents. TH Open. 2020 Oct;04(04):e288–99. https://doi. org/10.1055/s-0040-1718415
- 33. Ranucci M, Sitzia C, Baryshnikova E, Di Dedda U, Cardani R, Martelli F, et al. Covid-19-Associated Coagulopathy: Biomarkers of Thrombin Generation and Fibrinolysis Leading the Outcome. J Clin Med. 2020 Oct 28;9(11):3487. https://doi.org/10.3390/ jcm9113487
- 34. Fischer A, Sellner M, Mitusińska K, Bzywka M, Lill MA, Gyra A, et al. Computational selectivity assessment of protease inhibitors against sarscov-2. Int J Mol Sci. 2021 Feb 19;22(4):2065. https://doi.org/10.3390/ijms22042065
- 35. Venugopal A. Disseminated intravascular coagulation.IndianJAnaesth.2014Sep;58(5):603–8. https://dx.doi.org/10.4103%2F0019-5049.144666
- 36. Rosa A, Pye VE, Graham C, Muir L, Seow J, Ng KW, et al. SARS-CoV-2 can recruit a heme metabolite to evade antibody immunity Science Advances. 2021 May 1;7(22):eabg7607. https://doi.org/10.1126/ sciadv.abg7607
- Immenschuh S, Vijayan V, Janciauskiene S, Gueler F. Heme as a target for therapeutic interventions. Vol. 8, Frontiers in Pharmacology. Frontiers Research Foundation; 2017. Apr 4;8:146., https://

doi.org/10.3389/fphar.2017.00146

- Cavezzi A, Troiani E, Corrao S. COVID-19: Hemoglobin, Iron, and Hypoxia beyond Inflammation. A Narrative Review. Clin Pract. 2020 May 28;10(2):24–30. https://doi.org/10.4081/ cp.2020.1271
- 39. Maiti BK. Heme/Hemeoxygenase-1 System Is a Potential Therapeutic Intervention for COVID-19 Patients with Severe Complications. Vol. 3, ACS Pharmacology and Translational Science. American Chemical Society; 2020. p. 1032–4. https://doi.org/10.1021/acsptsci.0c00136
- 40. Wagener FA PPPSISAN. Targeting the heme-heme oxygenase system to prevent severe complications following COVID-19 infections. Antioxidants . 2020 Jun;9(6):540. https://doi.org/10.3390/ antiox9060540
- 41. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet (London, England). 2020 Feb;395(10223):497–506. https:// doi.org/10.1016/S0140-6736(20)30183-5
- 42. Marandino L, Necchi A, Aglietta M, Maio M Di. COVID-19 Emergency and the Need to Speed Up the Adoption of Electronic Patient-Reported Outcomes in Cancer Clinical Practice. JCO Oncol

Pract. 2020 Jun;16(6):295-298. doi: 10.1200/ OP.20.00237

- 43. Haleeqa MA, Alshamsi I, Al Habib A, Noshi M, Abdullah S, Kamour A, et al. Optimizing supportive care in COVID-19 patients: A multidisciplinary approach. Vol. 13, Journal of Multidisciplinary Healthcare. Dove Medical Press Ltd.; 2020. p. 877–80. doi: 10.2147/JMDH.S264168
- Zhou X, Cheng Z, Luo L, Zhu Y, Lin W, Ming Z, et al. Incidence and impact of disseminated intravascular coagulation in COVID-19 a systematic review and meta-analysis. Thromb Res. 2021 May 1;201:23– 9. https://doi.org/10.1016/j.thromres.2021.02.010
- 45. Lee KW, Khan AH, Ching SM, Devaraj NK, Baharin J, Chia PK, et al. Coronavirus Disease-2019: Knowledge and Practices Behaviour of Healthcare Workers at a University Teaching Hospital in Malaysia. Malaysian Journal of Medicine & Health Sciences. 2021:149-58
- 46. Ahmed N, Shakoor M, Vohra F, Abduljabbar T, Mariam Q, Rehman MA. Knowledge, Awareness and Practice of Health care Professionals amid SARS-CoV-2, Corona Virus Disease Outbreak. Pakistan J Med Sci. 2020 May;36(COVID19-S4):S49–56. https://dx.doi.org/10.12669%2Fpjms.36. COVID19-S4.2704